(Figures 1 and 2). Although the hydride ligands were not located, on the basis of steric grounds they are presumably located in the plane defined by P(3)-W-P(4), such that the overall coordination geometry is dodecahedral. Four-membered oxametallacycles, which are of particular interest as potential models for olefin oxidation,⁸ are rare compared with five- and six-membered derivatives.9 Furthermore, four-membered oxametallacycles derived from ortho metalation of aryloxy ligands are surprisingly rare,¹⁰ for which $W(PMe_3)_4H_2(\eta^2-OC_6H_4)$ represents a structurally characterized example.

More interesting situations arise in the reactions of the monosubstituted phenols, $2-RC_6H_4OH$ (R = CH₃, CH₂CH₃, CH- $(CH_3)_2$, $C(CH_3)_3$, in which a variety of potential C-H bond activation reactions are now possible, leading to the formation of either four-, five-, or six-membered oxametallacycles. For example, the reaction of 2-methylphenol gives the five-membered oxametallacycle W(PMe₃)₄H₂{ η^2 -OC₆H₃(CH₂)} as a result of selective sp³ C-H bond activation of the methyl substituent, in preference to the four-membered ortho-metalated alternative. However, in marked contrast, the corresponding reactions of the 2-ethyl-, 2-isopropyl-, and 2-tert-butylphenol derivatives give specifically the four-membered oxametallacycles as a result of selective sp^2 C-H bond activation at the ortho position.

The propensity for the formation of four-membered metallacycles within this system is striking, especially given the marked tendency of ortho-substituted tert-butyl groups in other systems to undergo facile metalation, with the resulting formation of six-membered oxametallacycles.9 Although at present we cannot address the question whether the formation of four-membered versus six-membered oxametallacycles in this system represents a kinetic or thermodynamic preference, we note that one contributing factor may be a consequence of the 18-electron configuration of the tungsten centers in these oxametallacycles. The six-membered oxametallacycle complexes that are derived from aryloxy ligands typically possess electron-deficient metal centers. Structural studies on these oxametallacycles demonstrate that formation of the six-membered ring allows for large M-O-C bond angles, thus enabling favorable lone-pair donation from oxygen to the electron-deficient metal center.⁹ Such lone-pair donation would clearly stabilize an oxametallacycle structure of an electron-deficient metal center. However, for the 18-electron complexes described here, lone-pair donation would not be expected to contribute significantly to oxametallacycle stability, and thus the preference for six-membered ring formation would be lessened.

Studies indicate that the four- and five-membered oxametallacycles are also highly reactive. For example, $W(PMe_3)_4H_2$ - $(\eta^2 - OC_6H_4)$ is hydrogenated rapidly by H₂ at room temperature to give W(PMe₃)₄H₃(OC₆H₅).¹¹ Deuterium labeling demon-

(6) Crystal data for W(PMe₃)₄H₂{ η^2 -OC₆H₂Me₂(CH₂)}: monoclinic, P2₁/n (No. 14), a = 9.898 (3) Å, b = 28.065 (9) Å, c = 10.663 (3) Å, $\beta = 104.33$ (2)°, V = 2870 (1) Å³, Z = 4, $\rho_{abled} = 1.45$ g cm⁻³, μ (Mo K α) = 44.9 cm⁻³ λ (Mo K α) = 0.71073 Å (graphite monochromator); 4474 unique reflections with 3° < 2 θ < 48° were collected of which 2556 reflections with $F > 6\sigma(F)$ were used in refinement; R = 0.0455, $R_w = 0.0451$, GOF = 1.274.

 $F > 6\sigma(F)$ were used in refinement; R = 0.0455, $R_w = 0.0451$, GOF = 1.274. (7) Examples of four-membered oxametallacycles include (AsPh₃)₂Pt-{OC(CN)₂C(CN)₂},^{7a.b} (PPh₃)₂Pt(CH₂OCH₂),^{7c} (η^5 -C₅Me₃)(2R(OCe₃)]₂,^{7l} and (η^5 -C₃Me₃)₂Zr(OCPh=CPh).^{7e} (a) Schlodder, R.; Ibers, J. A.; Lenarda, M.; Graziani, M. J. Am. Chem. Soc. 1974, 96, 6893-6900. (b) Lenarda, M.; Ros, R.; Traverso, O.; Pitts, W. D.; Baddley, W. H.; Graziani, M. Inorg. Chem. 1977, 16, 3178-3182. (c) Hoover, J. F.; Stryker, J. M. J. Am. Chem. Soc. 1989, 111, 6466-6468. (d) Klein, D. P.; Hayes, J. C.; Bergman, R. G. J. Am. Chem. Soc. 1988, 1/0, 3704-3706. (e) Ho, S. C.; Hentges, S.; Grubbs, R. H. Organometallics 1988, 7, 780-782. (f) Vaughan, G. A.; Hillhouse, G. L.; Lum, R. T.; Buchwald, S. L.; Rheingold, A. L. J. Am. Chem. Soc. 1988, 110, 7215-7217. (g) Vaughan, G. A.; Hillhouse, G. L.; Rheingold, A. L. J. (8) (a) Collman, J. P.; Hegedus, L. S.; Norton, J. R.; Finke, R. G. Prin-ciples and Applications of Organotransition Metal Chemistry; University

(a) Coliman, J. F., Hegedus, L. S., Holton, J. R., Finke, R. O. Principles and Applications of Organotransition Metal Chemistry; University Science Books: Mill Valley, CA, 1987.
(9) (a) Rothwell, I. P. Acc. Chem. Res. 1988, 21, 153–159. (b) Rothwell, I. P. Polyhedron 1985, 4, 177–200. (c) Yu, J. S.; Fanwick, P. E.; Rothwell, I. P. J. Am. Chem. Soc. 1990, 1/2, 8171–8172.

(10) The ortho-metalated ruthenium derivatives, $(PMe_3)_4Ru(\eta^2-OC_6H_4)$ and (PMe₃)₃Ru(η³-OC₄H₄), have recently been synthesized: Hartwig, J. F.; Andersen, R. A.; Bergman, R. G., personal communication. strates that this reaction proceeds via initial reductive elimination of the metallacycle-hydride unit (eq 2).

$$W(PMe_3)_4H_2(\eta^2 \cdot OC_6H_4) \rightarrow [W(PMe_3)_4H(OC_6H_5)] \xrightarrow{D_2} W(PMe_3)_4HD_2(OC_6H_5)] \xrightarrow{(D_2)} W(PMe_3)_4HD_2(OC_6H_5) (2)$$

In conclusion, these studies have demonstrated the selectivity with which electron-rich tungsten complexes may form four- and five-membered oxametallacycle complexes, in contrast to the more commonly observed six-membered derivatives that are obtained for electron-deficient metal complexes.

Supplementary Material Available: Tables of spectroscopic data for all new compounds, crystal and intensity collection data, atomic coordinates, bond distances and angles, and anisotropic displacement parameters and ORTEP drawings (21 pages); tables of observed and calculated structure factors for W(PMe₃)₄H₂(η^2 - OC_6H_4) and $W(PMe_3)_4H_2[\eta^2 - OC_6H_2Me_2(CH_2)]$ (28 pages). Ordering information is given on any current masthead page.

(11) Chiu, K. W.; Jones, R. A.; Wilkinson, G.; Galas, A. M. R.; Hursthouse, M. B.; Malik, K. M. A. J. Chem. Soc., Dalton Trans. 1981, 1204-1211.

Peptide Architecture. Design of Stable α -Helical Metallopeptides via a Novel Exchange-Inert Ru^{III} Complex

M. Reza Ghadiri* and A. Katharina Fernholz

Departments of Chemistry and Molecular Biology Research Institute of Scripps Clinic La Jolla, California 92037 Received August 14, 1990

There is considerable interest in the design of linear polypeptides that can adopt stable and well-defined solution conformations. Such systems can serve as models for the study of the early events in protein folding¹ as well as possess substantial utility in the design of peptide-based therapeutic agents.² However, in spite of a few encouraging results, design of short monomeric peptides with stable secondary structure conformations in water has not been forthcoming.³ Here we report for the first time the utility of an exchange-inert metal complex^{4,5} in the formation of remarkably stable α -helical peptides. Formation of a macrocyclic cis-[Ru- $(NH_3)_4L_2$ ³⁺ complex, where L_2 are the side chains of two histidines in positions i and i + 4 of a peptide, is shown to be a simple and effective method for constraining the intervening chain in an α -helical conformation and effecting helix nucleation. A 17residue polypeptide functionalized in this way has a melting

^{(1) (}a) Wright, P. E.; Dyson, H. J.; Lerner, R. A. Biochemistry 1988, 27, 7167. (b) Oas, T. G.; Kim, P. S. Nature 1988, 336, 42. (c) Baldwin, R. L. TIBS 1989, 14, 291. (d) Vasquez, M.; Pincus, M. R.; Scheraga, H. A. Biopolymers 1987, 26, 351.
(2) (a) Kaiser, E. T. TIBS 1987, 12, 305. (b) Taylor, J. W.; Kaiser, E. T. Pharmacol. Rev. 1986, 38, 291. (c) Finn, F. M.; Hofmann, K. Acc. Chem. Res. 1973, 6, 169. (d) Also see ref 3c.
(3) (a) Kemp, D. S.; Curran, T. P. Tetrahedron Lett. 1988, 29, 4931, 4935. (b) Arrhenius, T.; Chaing, L.-C.; Lerner, R. A.; Satterthwait, A. C. Vaccines 1989, 17. (c) Felix, A. M., et al. Int. J. Pept. Protein Res. 1988, 32, 441. (d) Kahn, M.; Wilke, S.; Chen, B.; Fujita, K. J. Am. Chem. Soc. 1988, 110, 1638. (e) Olson, G. L.; Voss, M. E.; Hill, D. E.; Kahn, M.; Madison, V. S.; Cook, C. M. J. Am. Chem. Soc. 1990, 112, 323. (f) Ravi, A.; Balaram, P. Tetrahedron 1984, 40, 2577. (g) Kemp, D. S.; Stites, W. E. Tetrahedron Lett. 1988, 29, 5057. (h) Nagai, U.; Sato, K. Tetrahedron Lett. 1985, 26, 647. (i) 1988, 29, 5057. (h) Nagai, U.; Sato, K. Tetrahedron Lett. 1985, 26, 647. (i) Feigel, M. J. Am. Chem. Soc. 1986, 108, 181.

⁽⁴⁾ For the earliest definition, see: Taube, H. Chem. Rev. 1952, 50, 69. (4) For the earliest definition, see: Taube, H. Chem. Rev. 1952, 50, 69.
(5) For the utility of exchange-inert metal ions in the study of biological macromolecules, see: (a) Van Wart, H. E. Methods Enzymol. 1988, 158, 95.
(b) Barton, J. K. Comments Inorg. Chem. 1985, 3, 321. (c) Metal-Ligand Interactions in Organic Chemistry and Biochemistry; Pullman, B., Goldblum, N., Eds.; D. Reidel: Boston, 1977. (d) Margalit, R.; Pecht, I.; Gray, H. B. J. Am. Chem. Soc. 1983, 105, 301. (e) Friedman, A. E.; Chambron, J.-C.; Sauvage, L.P.; Turro, N. L.; Barton, I. K. Lem. Chem. Soc. 1990, 1/2, 4960. Sauvage, J.-P.; Turro, N. J.; Barton, J. K. J. Am. Chem. Soc. 1990, 112, 4960.

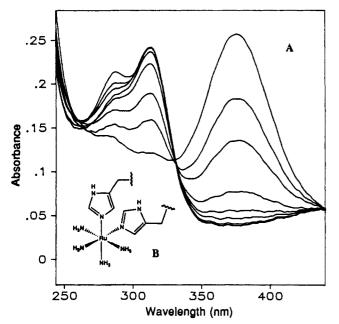


Figure 1. (A) Absorption spectra of tetraamineruthenium(III)-peptide 1 complex (6.0 \times 10⁻⁵ in water) at pH 4.7 (maxima at 313 nm), 5.5, 6.6, 7.1, 7.5, and 7.9 (maxima at 376 nm). (B) cis-[Ru(NH₃)₄(His₂peptide)]³⁺ chromophore.

temperature of 35 °C and exhibits 80% a-helicity at 21 °C.

In a recent study, we described the use of labile transition-metal complexes in the formation of α -helical peptides.⁶ In order to extend the scope and utility of this methodology to systems suitable for study under normal physiological conditions, peptide-metal complexes must be designed which are considerably more resistant to ligand exchange. However, unlike the exchange-labile ions which can be easily incorporated by virtue of their rapid exchange rates, formation of exchange-inert metallopeptide complexes is rather difficult.^{5a} In peptides and proteins where a number of reactive side-chain functionalities are present, chemoselectivity in the formation of an exchange-inert metal complex is the paramount issue and is primarily dependent on the careful choice of metal ion, its oxidation state, and the ligands employed.⁵ The high affinity of aquopentaammineruthenium(II) for surface-accessible histidine residues⁷ prompted us to examine the feasibility of forming stable macrocyclic cis-[Ru(NH₃)₄(His₂-peptide)]³ complexes of peptides 1 and 2 and further explore the utility of this approach in the induction and stabilization of the α -helical conformation. These peptides have, in addition to two histidines, two aspartic acid and two lysine residues in the relative i and i + 4 positions and are therefore good models to test the chemoselectivity of the complex-forming reaction. Peptides 1 and 2 were modified in the presence of cis- $[Ru(NH_3)_4(H_2O)_2]^{2+}$ in a degassed Tris buffer (50 mM, pH 7.1) for 6 h.8 The reaction mixture was air oxidized, and the metallopeptide was purified by either ion exchange or reversed-phase chromatography.9

peptide 1: acetyl-AEAAAKEAAAKHAAAHA-CONH₂

peptide 2: acetyl-AEAAAKHAAAHEAAAKA-CONH₂

(6) Ghadiri, M. R.; Choi, C. J. Am. Chem. Soc. 1990, 112, 1630.

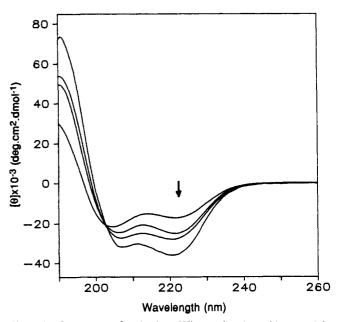


Figure 2. CD spectra of ruthenium(III)-complexed peptides 1 and 2. From top curve (arrow) to the bottom: (a) cis-Ru(NH₃)₄-peptide 2, 6.3 × 10⁻⁶ M in water, pH 6.0, at 20 °C and (b) at 0 °C; (c) cis-Ru-(NH₃)₄-peptide 1, 5.0 × 10⁻⁶ M in water, pH 6.1, at 20 °C and (d) at 0 °C.

Chemoselective functionalization of histidine residues is supported by NMR studies. The histidine C-2 and C-4 protons of peptide 2 occurring at δ 8.52 and 8.51 and at 7.20 and 7.19 undergo dramatic upfield shifts to δ 7.47 and 7.15 and at 6.98 and 6.83, respectively, upon attachment to a tetraammineruthenium(II) moiety. On the other hand, the histidine signals in the Ru(III)-peptide complex display paramagnetic shifting¹⁰ and appear as broad peaks at $\delta 0.56$ and -0.78^{11} Furthermore, the absorption spectrum of the Ru(III) complex of peptide 1 shows ligand to metal charge transfer bands at 287 and 313 nm below pH 6.5 which shift to 376 nm above pH 8 with an isosbestic point at 331 nm¹² (Figure 1). The pronounced shift in absorption maxima with increasing pH is consistent with the behavior of the Ru(III) ions containing coordinated imidazoles and is attributed to N-H deprotonation at the "pyrrole nitrogen".¹³ The absorption data give a pK_a value of 7.5 for the imidazole N-H of the coordinated histidine, which is 7 orders of magnitude lower than the pK_a of uncomplexed histidine.¹⁴ The spectral similarity with the simple ruthenium(III)-imidazole complexes and the characteristically facile metal-ion-promoted imidazole N-H ionization15 further support the formation of the desired complex.

(10) Bertini, I.; Luchinat, C. NMR of Paramagnetic Molecules in Bio-logical Systems; Benjamin-Cummings Publishing Co.: Menlo Park, 1986. (11) Similar paramagnetic shifts have been observed when His-33 of cy-

(1) Similar paramagnetic simils have been observed when His 35 of cy-tochrome c was functionalized with pentaammineruthenium(III). Gray, H. B; Meade, T. J., personal communication.
(12) Peptide 2 also exhibits similar UV spectra and pH profile.
(13) (a) Sundberg, R. J.; Bryan, R. F.; Taylor, I. F., Jr.; Taube, H. J. Am. Chem. Soc. 1974, 96, 381.
(b) Sundberg, R. J.; Gupta, G. Bioinorg. Chem. 1973, 3, 39.

(14) Uncomplexed imidazole deprotonates with a pK_a of around 14.5. Perrin, D. D. Dissociation Constants of Organic Bases in Aqueous Solution; Butterworth and Co. Ltd.: London, 1965; p 1393.

(15) For pK_a values of other transition-metal complexes of histidine, see: (a) Morris, P. J.; Martin, R. B. J. Am. Chem. Soc. **1970**, 92, 1543. (b) Pitner, T. P.; Wilson, E. W., Jr.; Martin, R. B. Inorg. Chem. **1972**, 11, 738. (c) Sundberg, R. J.; Martin, B. Chem. Rev. **1974**, 74, 471. (d) George, P.; Hanania, G. I.; Irvine, D. H.; Abu-Issa, I. J. Chem. Soc. **1964**, 5689.

⁽⁶⁾ Ghadiri, M. R.; Choi, C. J. Am. Chem. Soc. 1990, 112, 1630. (7) (a) Matthews, C. R.; Erickson, P. M.; Van Vliet, D. L.; Perersheim, M. J. Am. Chem. Soc. 1978. 100, 2260. (d) Recchina, J.; Matthews, C. R.; Rhee, M.-J.; Horrocks, W. D., Jr. Biochim. Biophys. Acta 1982, 702, 105. (c) Margalit, R.; Kostic, N. M.; Che, C.-M.; Blair, D. F.; Chiang, H.-J.; Pecht, l.; Shelton, J. B.; Shelton, J. R.; Schroeder, W. A.; Gray, H. B. Proc. Natl. Acad. Sci. U.S.A. 1984, 81, 6554. (d) Bowler, B. E.; Meade, T. J.; Mayo, S. L.; Richards, J. H.; Gray, H. B. J. Am. Chem. Soc. 1989, 111, 8757 and references cited therein. (e) Ratilla, E. M. A.; Brothers, H. M., II; Kostic, N. M. J. Am. Chem. Soc. 1987, 109, 4592. (8) cis-[Ru(NH₃)₄(H₂O)₂]²⁺ was freshly prepared by the reduction of cis-[Ru(NH₃)₄(Ll₂]Cl in the presence of zinc amalgam in a degassed Tris buffer, pH 7.0. For a convenient preparation of cis-[Ru(NH₃)₄Cl₂]Cl, see: Pell, S. D.; Sherban, M. M.; Tramontano, V.; Clarke, M. J. Inorg. Synth. 1989, 26, 65.

^{1989, 26, 65.}

⁽⁹⁾ Both ruthenium-peptide complexes are >98% pure as judged by analytical C₁₈RP-HPLC and cation-exchange chromatography (Polysulfoethyl Aspartamide HPLC, or BioRex-70 column). The total ruthenium content was Asparating to the period of t cating that the histidine residues are specifically modified. FAB MS of both peptides give rise to the expected molecular weight ion (m/z = 1797) as well as a prominent peak at 1729 corresponding to the loss of four NH₃ ligands.

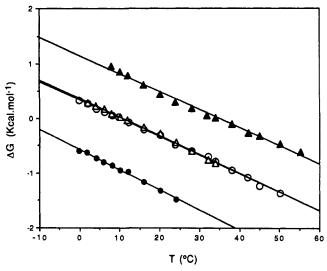


Figure 3. Plot of ΔG vs temperature for the unfolding of free and ruthenium-complexed peptides 1 and 2 [(5.0-6.3) × 10⁻⁶ M, in 20 mM MES, pH 6.0]: (•) peptide 2; (0) Ru(III)-peptide 2; (Δ) peptide 1; (Δ) Ru(111)-peptide 1.

Restriction of the conformational mobility of peptides 1 and 2 by the formation of an exchange-inert macrocyclic Ru(III) complex affords remarkably stable α -helical metallopeptides (Figure 2). The analysis of CD spectra¹⁶ of Ru(III)-complexed peptides 1 and 2 at 21 °C in water indicates 80% ($[\theta]_{222} = -28000$ deg·cm²·dmol⁻¹) and 50% ($[\theta]_{222} = -17300$ deg·cm²·dmol⁻¹) α -thelicity, respectively.¹⁷ In contrast, the free peptide 1 under similar conditions is 45% helical ($[\theta]_{222} = -16\,000 \text{ deg-cm}^2 \text{-dmol}^{-1}$) while uncomplexed peptide 2 exhibits the CD spectrum of a random coil structure.¹⁶ In order to assess differences in helix stability between the metal-ion-complexed and uncomplexed forms of peptides 1 and 2, the conformational stability of each peptide in the presence and absence of transition-metal ions was determined from heat denaturation studies.¹⁸ The analysis of the thermal denaturation curves afforded linear ΔG vs T and van't Hoff plots (Figure 3). The data suggest that the formation of the exchange-inert Ru(III) complex contributes up to 1 kcal·mol⁻¹ toward the stability of the α -helical conformation and dramatically increases the melting temperature of both peptides by about 25 °C. Complexed peptides 1 and 2 exhibit melting temperatures of 35 and 9.5 °C, respectively, while the corresponding free peptides have T_m values of 11 and -15.5 °C.

The above study unequivocally establishes that exchange-inert metal complexes can be effectively exploited in designing highly stable α -helical metallopeptides. The availability of a simple methodology for the formation of stable α -helical peptides can have considerable utility in the de novo design of biologically active peptides.

Stellwagen, E. Biochemistry 1990, 29, 894.

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Enantioselective Catalysis of the Triplex Diels-Alder **Reaction:** Addition of *trans*- β -Methylstyrene to 1,3-Cyclohexadiene Photosensitized with (-)-1,1'-Bis(2,4-dicyanonaphthalene)

Ji-In Kim and Gary B. Schuster*

Department of Chemistry, Roger Adams Laboratory University of Illinois, Urbana, Illinois 61801 Received August 22, 1990 Revised Manuscript Received October 11, 1990

The discovery of enantioselective photochemical sensitization reactions has been pursued with modest success since 1965 when Hammond and Cole reported that triplet energy transfer from an optically active amide gives a 7.7% enantiomeric excess (ee) of trans-1,2-diphenylcyclopropanes.¹ Similarly, Takamuku and co-workers observe modest enrichments from isomerization of trans-cyclooctenes sensitized by optically active esters.² Inoue and co-workers described an unusual temperature dependence for this reaction.³ The differential recognition of excited molecules bound to chiral surfaces by optically active quenchers was recently reported by Avnir and co-workers.⁴ Despite a massive effort to develop chiral Lewis acids that will induce optical activity in cycloadducts formed by the Diels-Alder reaction,⁵ only Vondenhof and Mattay have considered a photochemical route to this goal.⁶ They describe use of an optically active sensitizer in a radical cation Diels-Alder reaction, but do not report an enantiomeric excess in the cycloadducts that result.

We recently described the [4 + 2] cycloaddition of electron-rich dienes to electron-rich dienophiles in nonpolar solvents catalyzed by irradiation of electron-deficient arene sensitizers.⁷ This process

(5) (a) Helmchen, G.; Karge, R.; Weetman, J. In Modern Synthetic Methods; Scheffold, R., Ed.; Springer-Verlag: Berlin, 1986; Vol. 4, p 261. (b) Nachr. Chem., Tech. Lab. 1987, 35, 836. (c) Oppolzer, W. Angew. Chem., (d), Holm Nathala, S. (d), Yamamoto, H.; Maruoka, K.; Furata, K.;
Ikeda, N.; Mori, A. In Stereochemistry of Organic and Bioorganic Transformations; Bartmann, W., Sharpless, K. B., Eds.; VCH Publishers: Weinheim, 1987; p 13. (e) Maruoka, K.; Itoh, T.; Shirasaka, T.; Yamamoto, H. J. Am. Chem. Soc. 1988, 110, 310. (f) Evans, D. A.; Chapman, K. T.; Bisaha, J. J. Am. Chem. Soc. 1988, 110, 1238. (g) Narasaka, K.; Inoue, M.; Okada, N. Chem. Lett. 1986, 1109. (h) Chapuis, C.; Jurczak, J. Helv. Chim. Acta 1987, 70, 436. (i) Furuta, K.; Miwa, Y.; Iwanaga, K.; Yamamoto, H. J. Am. Chem. Soc. 1988, 110, 6254. (j) Paquette, L. E. In Asymmetric Synthesis; Morrison, J. D., Ed.; Academic: New York, 1984; Vol. 3, Chapter (k) Wasawa, N.; Sugimori, J.; Kawase, Y.; Narasaka, K. Chem. Lett. 1989, 1947. (l) Cossu, S.; Delogu, G.; De Lucchi, O.; Fabbri, D.; Licini, G. Angew. Chem., Int. Ed. Engl. 1989, 28, 766. (m) Narasaka, K.; Iwasawa, N.; Inoue, M.; Yamada, T.; Nakashima, M.; Sugimori, J. J. Am. Chem. Soc. 1989, 111, 5340. (n) Corey, E. J.; Imwinkelried, R.; Pikul, S.; Xiang, Y. B. J. Am. Chem. Soc. 1989, 111, 5493. (o) Kaufmann, D.; Boese, R. Angew. Chem., Int. Ed. Engl. 1990, 29, 545. Int. Ed. Engl. 1984, 23, 876. (d) Yamamoto, H.; Maruoka, K.; Furata, K.; Engl. 1990, 29, 545

(a) Condenhof, M.; Mattay, J. Tetrahedron Lett. 1990, 31, 985.
(b) Condenhof, M.; Mattay, J. Tetrahedron Lett. 1990, 31, 985.
(c) Calhoun, G. C.; Schuster, G. B. J. Am. Chem. Soc. 1984, 106, 6870.
(c) Calhoun, G. C.; Schuster, G. B. J. Am. Chem. Soc. 1986, 108, 8021.
(d) Calhoun, G. C.; Schuster, G. B. J. Am. Chem. Soc. 1986, 108, 8021. Akulbut, N.; Schuster, G. B. Tetrahedron Lett. 1988, 29, 5125. (e) Hartsough, D.; Schuster, G. B. J. Org. Chem. 1989, 54, 3. (f) Akbulut, N.;
 Hartsough, D.; Kim, J.-I.; Schuster, G. B. J. Org. Chem. 1989, 54, 2549.

^{(16) (}a) Woody, R. W. In The Peptides; Udenfriend, S., Meienhofer, J., Eds.; Academic Press Inc.: New York, 1985; Vol. 7, p 15. (b) Greenfield, N.; Fasman, G. D. *Biochemistry* **1969**, *8*, 4108. (c) Johnson, W. C., Jr. *Annu. Rev. Biophys. Chem.* **1988**, *17*, 145.

⁽¹⁷⁾ Ruthenium(11)-peptide complexes are monomeric in solution as inferred from the absence of any concentration dependency in the CD spectra in the measured range of $0.5-300 \,\mu$ M. The ruthenium complex does not make a significant contribution to the CD spectrum as judged by comparing the $[\theta]_{222}$ of heat and guanidine hydrochloride denatured ruthenium-peptide complexes to the $[\theta]_{222}$ of uncomplexed peptides under similar conditions. Peptide concentrations were determined by quantitative amino acid analysis Peptide concentrations were determined by quantitative amino acid analysis (average of three runs) using norleucine as internal standard. All reported CD data have an uncertainty of $\pm 2-5\%$. For 100% helix, $[\theta]_{222} = -35000$ and is based on several CD measurements for both peptides in various THF/H₂O mixtures at -10 to 20 °C. For 0% helix, an average value of $[\theta]_{222}$ = -5000 was determined from thermal denaturation curves in the range of 80-90 °C. Also see ref 6, and see: (a) Lyu, P. C.; Marky, L. A.; Kallenbach, N. R. J. Am. Chem. Soc. 1989, 111, 2733. (b) Shoemaker, K. R.; Kim, P. S.; York, E. J.; Stewart, J. M.; Baldwin, R. L. Nature 1987, 326, 563. (18) (a) Pace, N. C.; Shirley, B. A.; Thomson, J. A. In Protein Structure; Creighton, T. E., Ed.; IRL Press: Oxford, 1989; p 311. (b) Merutka, G.; Stellwagen, E. Biochemistry 1990, 29, 894.

⁽¹⁾ Hammond, G. S.; Cole, R. S. J. Am. Chem. Soc. 1965, 87, 3256. (1) Hammond, G. S.; Cole, K. S. J. Am. Chem. Soc. 1965, 87, 3256.
 (2) Inoue, Y.; Kunitomi, Y.; Takamuku, S.; Sakurai, H. J. Chem. Soc., Chem. Commun. 1978, 1024. Goto, S.; Takamuku, S.; Sakurai, H.; Inoue, Y.; Hakushi, T. J. Chem. Soc., Perkin Trans. 2 1980, 1678.
 (3) Inoue, Y.; Yokoyama, T.; Yamasaki, N.; Tai, A. J. Am. Chem. Soc. 1989, 111, 6480. Inoue, Y.; Yokoyama, T.; Yamasaki, N.; Tai, A. Mature 1989, 341, 225.

⁽⁴⁾ Avnir, D.; Wellner, E.; Ottolenghi, M. J. Am. Chem. Soc. 1989, 111, 2001